

Polymerase Chain Reaction (PCR) Utilizing Hot Start *Taq* DNA Polymerase

Introduction

Polymerase Chain Reaction (PCR) is a powerful technique used to amplify DNA through the use of the enzyme *Taq* DNA Polymerase. GoldBio Hot Start *Taq* DNA Polymerase is a thermostable DNA polymerase that possesses a 5'→3' polymerase activity and a 5' flap endonuclease activity. Hot Start *Taq* DNA Polymerase has been chemically modified to completely inactivate enzymatic activity until the initial heat activation step at the beginning of the PCR cycle. Hot Start PCR reduces non-specific amplification during the setup of the reaction and helps increase PCR specificity and sensitivity. This product is supplied with a 10x PCR reaction buffer containing MgCl₂, which produces a final Mg²⁺ concentration of 1.5mM. GoldBio *Taq* DNA Polymerase can be used in many applications including routine PCR, primer extension, colony PCR, and amplification of high GC content DNA with the 5x GC enhancer. Here, we describe a general protocol for the use of GoldBio Hot Start *Taq* DNA Polymerase.

Materials

- Hot Start *Taq* DNA Polymerase (GoldBio Catalog # [T-510](#) or [T-511](#))
- 10x PCR Buffer with Mg²⁺
- 5x GC enhancer
- 10mM dNTP (not supplied with T-510)

Not supplied:

- Primers
- H₂O PCR Grade

Note: Hot Start *Taq* DNA Polymerase should be stored at -20°C. The storage buffer contains 50mM Tris-HCl, 50mM KCl, 1mM DTT, 0.1mM EDTA, 50% glycerol, pH 7.5 at 25°C.

Note: 10x PCR Buffer with Mg²⁺ contains 100mM Tris-HCl pH 9.0, 15mM MgCl₂, 100mM KCl, 80mM (NH₄)₂SO₄, 0.5% Igepal CA 630.

Note: One unit is defined as the amount of enzyme that incorporates 10nmoles of dNTP into acid-insoluble form in 30 minutes at 72°C.

Method

1. Thaw the 10x PCR buffer, dNTPs, primers, and 5x GC enhancer (optional) on ice, and mix thoroughly.

2. Prepare a reaction mix for a 100 μ l reaction according to Table 1.

Note: The reaction mix contains all the components needed for the PCR reaction, except the template DNA.

Table 1. PCR reaction set up.

Component	100 μ l reaction
Template DNA	x μ l (0.01-0.5 μ g)
10x PCR Buffer	10.0 μ l
dNTP (10mM)	2.0 μ l
Forward primer	x μ l (0.1-0.5 μ M)
Reverse primer	x μ l (0.1-0.5 μ M)
5x GC enhancer (optional)	20.0 μ l
Hot Start <i>Taq</i> DNA Polymerase (5 U/ μ l)	0.5 μ l
H ₂ O	up to 100.0 μ l

3. Mix the reaction thoroughly.
4. Add template DNA to the individual PCR tubes containing the reaction mixture.

Note: Prepare the reaction mix on ice and immediately place reactions on thermocycler for PCR completion.

Note: Mix gently and spin briefly if necessary to collect the whole volume at the bottom of the tube.

5. Program the thermal cycler according to the manufacturer's instructions using a PCR cycling program similar to the program described in Table 2.

Note: The PCR program must start with an initial heat-activation step at 95°C for 15 minutes.

Table 2. Sample PCR cycling conditions.

Steps	Temp.	Time	Cycles
Initial Denaturation	95°C	15 min	1
Denaturation	94°C	30-60 sec	25-35
Annealing	52-66°C	30-60 sec	
Extension	72°C	1-2 min	
Final Extension	72°C	10 min	1
Hold	4-12°C		∞

- Place the PCR tubes in the thermal cycler and complete the cycling program.

Associated Products

- [dNTP mix \(GoldBio Catalog # D-900\)](#)
- [Hot Start Taq 2x Master Mix – 50 µl reaction \(GoldBio Catalog # T-512\)](#)
- [Hot Start Taq 2x Master Mix – 20 µl reaction \(GoldBio Catalog # T-513\)](#)
- [Taq DNA Polymerase \(GoldBio Catalog # T-514\)](#)
- [Taq DNA Polymerase plus dNTP \(GoldBio Catalog # T-515\)](#)
- [Taq DNA Polymerase with Dye \(GoldBio Catalog # T-516\)](#)
- [Taq DNA Polymerase with Dye plus dNTP \(GoldBio Catalog # T-517\)](#)
- [Taq DNA Polymerase 2x Premix with Dye \(GoldBio Catalog # T-518\)](#)
- [Hot Start Pfu DNA Polymerase \(GoldBio Catalog # P-650\)](#)
- [Hot Start Pfu DNA Polymerase plus dNTP \(GoldBio Catalog # P-655\)](#)
- [Pfu 2x DNA Polymerase Master Mix \(GoldBio Catalog # P-660\)](#)
- [Pfu DNA Polymerase \(GoldBio Catalog # P-665\)](#)
- [Pfu DNA Polymerase plus dNTP \(GoldBio Catalog # P-690\)](#)

References

- Chien, A., Edgar, D. B., & Trela, J. M. (1976). Deoxyribonucleic acid polymerase from the extreme thermophile *Thermus aquaticus*. *The Journal of Bacteriology*, 127, 1550-1557. Doi:10.1126/science.7683443.
- Lawyer, F. C., Stoffel, S., Saiki, R. K., Chang, S. Y., Landre, P. A., Abramson, R. D., & Gelfand, D. H. (1993). High-level expression, purification, and enzymatic characterization of full-length *Thermus aquaticus* DNA polymerase and a truncated form deficient in 5' to 3' exonuclease activity. *Genome Research*, 2(4), 275-287. Doi:10.1101/gr.2.4.275.
- Longley, M. J., Bennett, S. E., & Mosbaugh, D. W. (1990). Characterization of the 5' to 3' exonuclease associated with *Thermus aquaticus* DNA polymerase. *Nucleic Acids Research*, 18(24), 7317-7322. Doi:10.1093/nar/18.24.7317.
- Lyamichev, V., Brow, M., & Dahlberg, J. (1993). Structure-specific endonucleolytic cleavage of nucleic acids by eubacterial DNA polymerases. *Science*, 260(5109), 778-783. Doi:10.1126/science.7683443.